

---

# INTERNATIONAL JOURNAL OF CURRENT RESEARCH IN CHEMISTRY AND PHARMACEUTICAL SCIENCES

(p-ISSN: 2348-5213; e-ISSN: 2348-5221)  
www.ijrcrps.com

---



Research Article

## DESIGN AND DEVELOPMENT OF ORAL *IN SITU* GELS OF ACYCLOVIR

G.RAJESHWARI, DR.S.K.UMADEVI, JAYA SINGH, B.KASTURI BAI, MANISHA LACHORIYA,  
SAI VARDHAN

Department of Pharmaceutics, St.Pauls College of Pharmacy, Turkayamjal, Hyderabad, India.

Corresponding Author: drumadeviphd@gmail.com

---

### Abstract

The present investigation deals with the development and optimization of an *In situ* gelling formulation of Acyclovir. It is an antiviral drug. *In situ* gels are capable of releasing drug in a sustained manner affording relatively constant plasma profiles. The study evaluates the potential of oral sustained delivery of Acyclovir with sodium alginate and xanthan gum as gelling agents. Calcium ions present in the formulation helps in formation of gel when administered oral aqueous solutions. Aqueous solutions of sodium alginate and xanthan gum form gels on warming to body temperature, in the presence of cations. Different solutions (sols) were prepared varying the concentrations from 0.5%w/v to 2%w/v using sodium alginate and xanthan gum. The sols were evaluated for rheological properties using brookfield viscometer. *In vitro* release studies were carried out using simulated gastric fluid (p<sup>H</sup> 1.2) for 2 hrs followed by phosphate buffer (p<sup>H</sup> 6.8) for 6 hrs respectively and cumulative amount of drug release was analyzed by UV spectrophotometry. The optimized formulation was able to release its contents up to 8 hrs. These *In situ* gels are, thus suitable for oral sustained release of Acyclovir.

**Keywords:** Acyclovir, *In situ* gels, sodium alginate, xanthan gum, sustained drug delivery.

---

### Introduction

The antiviral drugs are mostly designed to suppress the replication of HIV, herpes viruses, the hepatitis B and C viruses, and influenza A and B viruses rather than killing them. cyclovir is a guanosine analog antiviral drug that acts as an antimetabolite. It is used to treat herpes simplex viral infections, varicella zoster(chicken pox) and herpes zoster( shingels). They inhibits viral DNA polymerase competitivelyand by termination of DNA synthesis by incorporation into DNA.

It is difficult to design safe and effective dosage forms of antiviral drugs because viruses use the host's cells to replicate. So it is hard to find targets for the drug that would interfere with the virus without also attacking the host organism's cells. Herpes simplex

virus 1 and 2 (HSV-1 and HSV-2), also known as human herpes virus 1 and 2 (HHV-1 and HHV-2), are two members of the herpes virus family, Herpesviridae, that infect humans. Both HSV-1 (which produces most cold sores) and HSV-2 (which produces most genital herpes) are ubiquitous and contagious.

The antiviral drugs are thus interfere with viral replication, and suppress the physical severity of outbreak-associated lesions, and minimize the transmission to others. Studies of vulnerable patient populations have revealed that daily use of antivirals such as Acyclovir and Valacyclovir can reduce reactivation rates.

Treatment in Children: it decreases symptoms if oral Acyclovir is started within 24 hours of onset of symptoms. Use of acyclovir therefore is not currently recommended for immunocompetent individuals (i.e., otherwise healthy persons without known immunodeficiency or on immunosuppressive medication). It is not prescribed for Children younger than 12 years old and older than one month.

In adults treatment with antiviral drugs (e.g. Acyclovir or valacyclovir) are generally prescribed, as long as it is started within 24–48 hours from the onset symptoms. Antiviral medicines do not kill the virus, but stop it from multiplying<sup>1</sup>.

An ample number of research works have been exploited in the development of *In situ* gelling systems. The development of *In situ* gels has attracted more attention over the past few decades. *In situ* gels are delivery systems which are capable of releasing drug in a sustained manner to maintain a constant plasma concentrations of drugs. These hydro gels are solutions at room temperature which undergo gelation when in contact with body fluids or change in pH. The gelation may occur due to changes in temperature, pH and ionic content<sup>1</sup>. *In situ* gels are drug delivery systems which possess potential advantages like simple manufacturing process, ease of administration, reduced frequency of administration, and improved patient compliance.<sup>2-4</sup>

*In situ* gels are resembling mucoadhesive drug delivery system in their mechanism of absorption in the GI tract. They swell to form a strong gel that is capable of prolonging the residence time of the drugs on the walls of GI tract. *In situ* gels can be administered by various routes such as oral<sup>8</sup>, ocular<sup>9</sup>, rectal<sup>10</sup>, vaginal<sup>11</sup>, injectable<sup>12</sup> and intra-peritoneal routes<sup>12</sup>.

In the present work, two natural bio-degradable polymers namely sodium alginate and xanthan gum are used for this purpose at various concentrations. As a part of this study effect of polymer combination on pattern and duration of *In vitro* dissolution study was also carried out. The drug dissolution data were fitted with various kinetic equations such as zero order, first order, korsmeyer peppas, hixson-crowell and higuchi to find the best fit model for the drug release.

## Materials and Methods

### Materials

The drug Acyclovir was procured as gift sample from Orex pharma, Dombivali, India, Sodium alginate and xanthan gum were procured from SD fine chemicals, India. All other chemicals used in this study were of analytical grade.

### Methods

**Table 1. Acyclovir *In situ* gels with xanthan gum and sodium alginate (F1-F8)**

| Sr. no | Formulations | Acyclovir (mg) | Xanthan gum (mg) | Sodium alginate (mg) | Sodium citrate (%w/v) | Calcium Chloride (%w/v) |
|--------|--------------|----------------|------------------|----------------------|-----------------------|-------------------------|
| 1      | F1           | 1              | 0.5              | -                    | 0.17                  | 0.016                   |
| 2      | F2           | 1              | 1                | -                    | 0.17                  | 0.016                   |
| 3      | F3           | 1              | 1.5              | -                    | 0.17                  | 0.016                   |
| 4      | F4           | 1              | 2                | -                    | 0.17                  | 0.016                   |
| 5      | F5           | 1              | -                | 0.5                  | 0.25                  | 0.075                   |
| 6      | F6           | 1              | -                | 1                    | 0.25                  | 0.075                   |
| 7      | F7           | 1              | -                | 1.5                  | 0.25                  | 0.075                   |
| 8      | F8           | 1              | -                | 2                    | 0.25                  | 0.075                   |

### Compatibility Studies

#### Fourier transform infra-red spectroscopy (FTIR)

Drug and polymer compatibility studies were carried out using FTIR spectral studies to establish the

possible interaction in the formulations. Samples about 5 mg was mixed thoroughly with 100 mg potassium bromide IR powder and compacted under vacuum at a pressure of about 12 Psi for 3 minutes and then the FTIR spectra were recorded between 400 and 4000 cm<sup>-1</sup>.

**Preparation of *in situ* gels of acyclovir****Preparation of xanthan gum *In situ* gels**

Xanthan gum solutions of concentrations 0.5 -2.0 % w/v were prepared by adding the gum to ultrapure water containing 0.17 % w/v sodium citrate and heated to 90°C while stirring. After cooling to below 40°C appropriate amounts of calcium chloride 0.016 % w/v was added while stirring. Acyclovir (50 mg) was then dissolved in 0.1N HCl solution and added slowly to the above gum solution while stirring on a magnetic stirrer so that there was proper and homogeneous dispersion of the drug. The HCl was neutralized with 0.1N NaOH.

**Preparation of sodium alginate *In situ* gels**

Sodium alginate suspensions were also prepared in a similar manner. Sodium alginate solutions of concentrations 0.5-2.0% w/v were prepared by adding the alginate to ultrapure water containing 0.25% w/v sodium citrate and heating to 60°C. After cooling to below 40°C appropriate amounts of calcium chloride 0.075 % w/v was added while stirring. Acyclovir (50mg) was then dissolved in 0.1N HCl solution and added slowly to the above alginate solution while stirring on a magnetic stirrer so that there was proper and homogeneous dispersion of the drug. The HCl was neutralized with addition of 0.1N NaOH.(Table 1).

**Evaluation of *In situ* gels**

The prepared *In situ* gels were evaluated for the following parameters.

- A) Drug content
- B) Rheological properties
- C) *In vitro* drug release study
- D) *In vitro* drug release kinetics.

**A) Drug content**

*In situ* gels of Acyclovir equivalent to 50mg was taken and dissolved in 10ml of 0.1N HCl buffer for 1 hr. Then that solution was filtered. From that 1ml is taken and diluted to 10ml and filtrate was analysed by UV Spectrophotometer at 255nm.

**B) Rheological properties of Sols and Gels**

The viscosity of the sols and gels were determined at ambient condition using Brookfield viscometer using 2 ml aliquot of the sample.

**C) *In vitro* drug release study**

The drug release rate from gels were determined by using USP dissolution apparatus Type I (basket-type). A weighed amount of gels equivalent to 50 mg of drug (Acyclovir) was weighed and placed in the basket. Dissolution medium used was 0.1 N HCl (pH 1.2, 900 ml) for first 2 hours and maintained at  $37 \pm 0.5^\circ\text{C}$  at a rotation speed of 100 rpm. 5 ml of sample was withdrawn at each 30mins interval, later this interval was extended to 2 hrs. Sample was then passed through a filter, and analyzed spectro photometrically at 255nm to determine the concentration of Acyclovir present in the dissolution medium. The initial volume of dissolution medium was maintained by adding 5 ml of fresh dissolution medium after each withdrawal. The dissolution study was continued using phosphate buffer (pH  $6.8 \pm 1$ , 900ml) for next 6 hrs. The cumulative % drug release was calculated using standard calibration curve.

**D) IN VITRO DRUG RELEASE KINETICS****Zero order kinetics:**

Drug dissolution from dosage forms that do not disaggregate and release the drug slowly can be represented by the equation:

$$Q_0 - Q_t = K_0 t$$

Where,  $Q_t$  is the amount of drug dissolved in time  $t$ ,  $Q_0$  is the initial amount of drug in the solution (most times,  $Q_0 = 0$ )

$K_0$  is the zero order release constant expressed in units of concentration/time.

When the data is plotted as cumulative percent drug release versus time, if the plot is linear then the data obeys zero-order release kinetics, with slope equal to  $K_0$ .

**First order kinetics**

The release of the drug which followed first order kinetics can be expressed by the equation:

$$\log C = \log C_0 - K t / 2.303$$

Where,  $C_0$  is the initial concentration of drug,  
 $K$  is the first order rate constant  
 $t$  is the time.

The data obtained are plotted as log cumulative percentage of drug remaining vs time which would yield a straight line with a slope of  $-K/2.303$ .

**Higuchi model:**

The release of the drug which follows higuchi kinetics can be expressed by the equation:

$$Q = K_H * t^{1/2}$$

Where,  $K_H$  is the Higuchi dissolution constant  
 $Q$  is the amount of drug released in time  $t$

The data obtained were plotted as cumulative percentage drug release versus square root of time.

**Hixson Crowell model:**

Hixson Crowell recognized that the particle regular area is proportional to the cube root of its volume. Hixson Crowell equation is

$$W_o^{1/3} - W_t^{1/3} = K_s * t$$

Where,  
 $W_o$ - initial amount of drug in pharmaceutical dosage form  
 $W_t$ - remaining amount of drug in pharmaceutical dosage form at time  $t$   
 $K_s$ -constant incorporating the surface volume relation.

The data obtained are plotted as (percent drug release)  $1/3$  verses time.

**Korsmeyer-Peppas model:**

To find out the mechanism of drug release, drug release data were fitted in Korsmeyer-Peppas equation which is expressed as:

$$Q/Q_0 = k t^n$$

Where,  $K_0$  to  $K_2$  were release rate constants  
 $Q/Q_0$  was fraction of drug released at time  $t$ ,

$K$  was constant and  $n$  was diffusion constant that indicates general operating release mechanism. For Fickian (diffusion controlled)  $n = 0.5$ ; for non Fickian (anomalous/zero order) release 'n' value is in between 0.5 to 1.0; for zero order release  $n=1.0$ ; for super case transport II,  $n > 1.0$ .

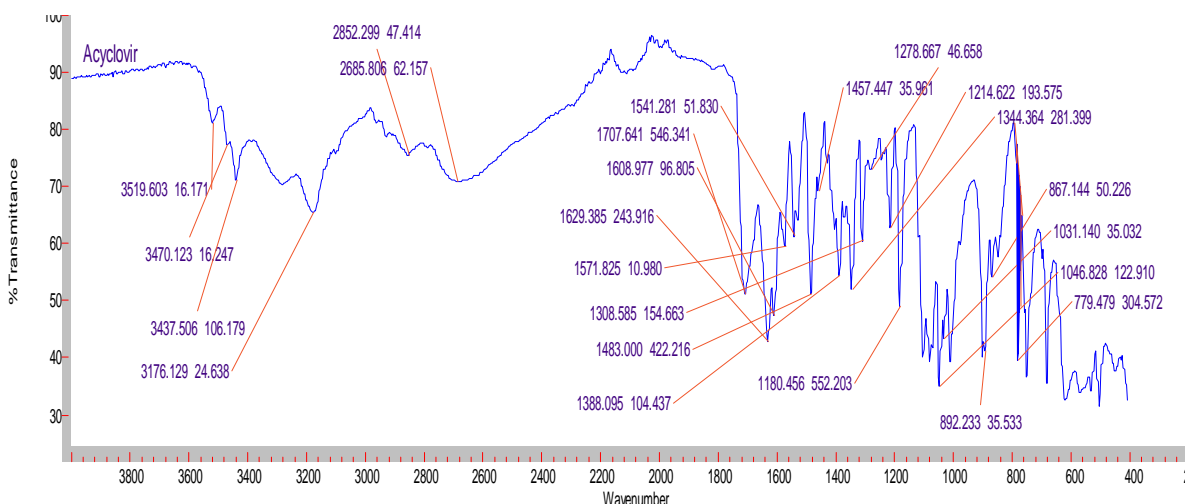
To study the release kinetics, data obtained from *In vitro* drug release studies were plotted as log cumulative percentage drug release versus log time.

**Results and Discussion**

**PREFORMULATION STUDIES**

**Compatibility Studies**

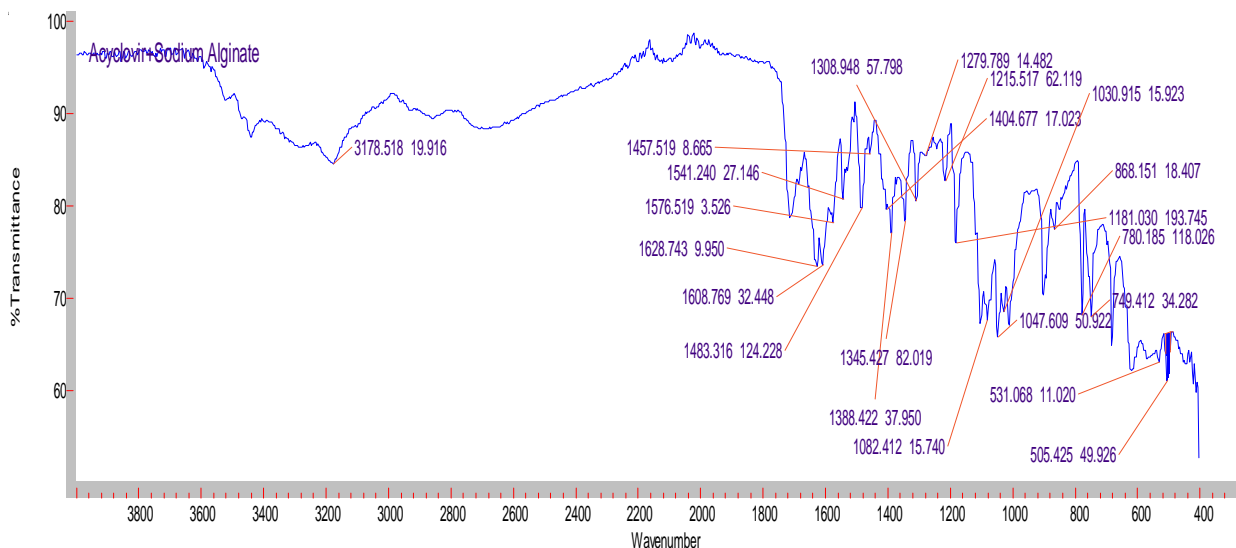
Drug and polymer compatibility studies were carried out using FTIR spectral studies to establish the possible interaction in the formulations. The IR spectrum of Acyclovir, xanthan gum, sodium alginate & their physical mixture is shown in Fig .1-3.



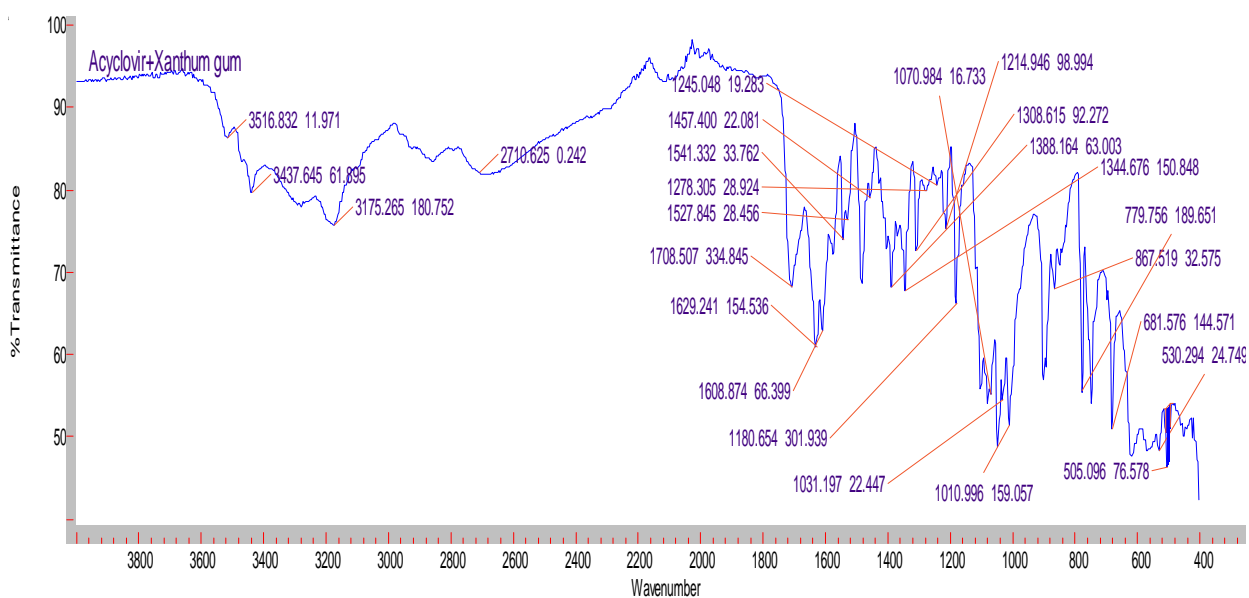
**Fig.1 FT-IR spectrum of pure drug acyclovir**

**Table 2. Characteristic peaks of Acyclovir**

| Sr.No | Functional group | Theoretical value | Peak wave values of pure drug (Cm <sup>-1</sup> ) |
|-------|------------------|-------------------|---|
| 1     | C=N              | 1670-1780         | 1707  |
| 2     | NH               | 3300-3500         | 3438  |
| 3     | C=O              | 1670-1780         | 1708  |
| 4     | C-O              | 1050-1150         | 1082  |
| 5     | C-N              | 1030-1230         | 1030  |



**Fig .2 FT-IR Spectrum of acyclovir + sodium alginate**



**Fig.3 FT-IR Spectrum of acyclovir + xanthan gum**

**Table 3. Comparative FTIR Interpretation of Acyclovir with Excipients**

| Sl.No | Functional Group | Theoretical value | pure drug(Cm <sup>-1</sup> ) | Pure drug+sodium alginate | Pure drug +xanthan gum |
|-------|------------------|-------------------|------------------------------|---------------------------|------------------------|
| 1     | -C=N             | 1670-1780         | 1707                         | 1686                      | 1708                   |
| 2     | -NH              | 3300-3500         | 3438                         | 3436                      | 3470                   |
| 3     | -C=O             | 1670-1780         | 1708                         | 1686                      | 1708                   |

From the above table 2 the peak values of mixture of drug with excipients is within the range of peak values of pure drug. This implies that the excipients are compatible with the drug since their combination did not alter the functional groups of pure drug.

#### Drug content determination

*In situ* gel of Acyclovir equivalent to 50mg was taken and dissolved in 10ml of 0.1N HCl buffer for 1 hr and then that solution was filtered. From that 1ml is taken diluted to 10ml and filtrate was analysed by UV Spectrophotometer at 255 nm.

**Table 4: Drug Content determination of Acyclovir *In situ* gels with Xanthan gum and sodium alginate F1-F8**

| FORMULATION | DRUG CONTENT (%)±SD |
|-------------|---------------------|
| F1          | 70±0.62             |
| F2          | 73±0.85             |
| F3          | 69.7±0.60           |
| F4          | 68.9±0.58           |
| F5          | 71.34±0.72          |
| <b>F6</b>   | <b>95.2±0.40</b>    |
| F7          | 84.2±0.92           |
| F8          | 79.2±0.54           |

UV spectroscopic analysis of one ml of all the formulation diluted with 0.1N HCl revealed that all the

formulations contains from 68.9±0.58(F4) to **95.2±0.40** %(F6) of the drug the results are triplicate of analysis.

#### Rheological properties

**Table 5. Rheological properties of acyclovir *In situ* gels**

| Formulation Code | Drug : polymer | Concentration (%) | Speed (RPM) | Viscosity (CP) |
|------------------|----------------|-------------------|-------------|----------------|
| F1               | 1:0.5          | 0.5               | 150         | 260            |
| F2               | 1:1.0          | 1                 | 150         | 325            |
| F3               | 1:1.5          | 1.5               | 150         | 350            |
| F4               | 1:2            | 2                 | 150         | 370            |
| F5               | 1:0.5          | 0.5               | 150         | 175            |
| <b>F6</b>        | <b>1:1.0</b>   | <b>1</b>          | <b>150</b>  | <b>255</b>     |
| F7               | 1:1.5          | 1.5               | 150         | 272            |
| F8               | 1:2            | 2                 | 150         | 270            |

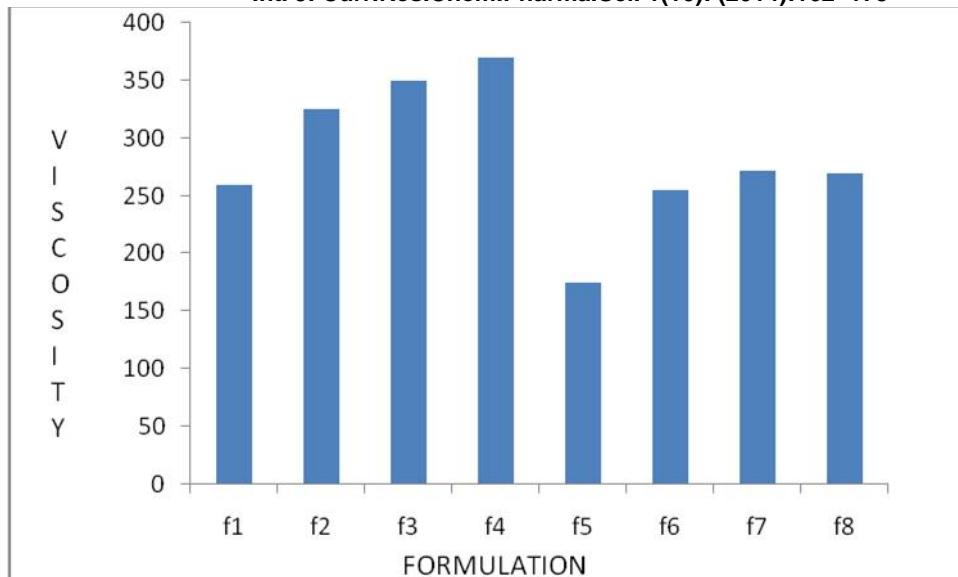


Fig .4. viscosity of acyclovir *In situ* gels (F1-F8) at 150 RPM

**Dissolution studies of *in situ* gels**

The *In vitro* drug release studies were carried out by using USP type I dissolution apparatus. The *In situ* gels were placed in a basket. The dissolution medium used was 900ml of 1.2 pH buffers at 37°C for first 2 hours followed by 6.8 pH phosphate buffers at 37°C for the rest of the study.

The cumulative present drug release of F1 to F8 formulations at various time intervals was calculated. The cumulative percent drug release in all formulations was plotted against time in fig.5-8. The maximum percent of drug release was found in F6 formulation.

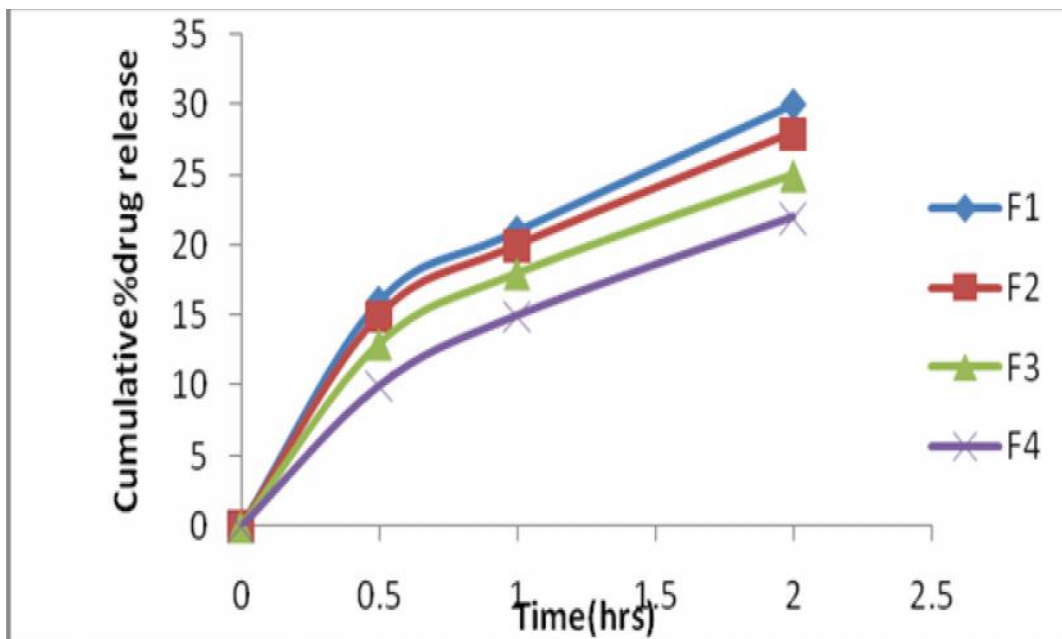


Fig. 5 *In vitro* cumulative% drug release of Acyclovir *In situ* gels with xanthan gum in pH 1.2

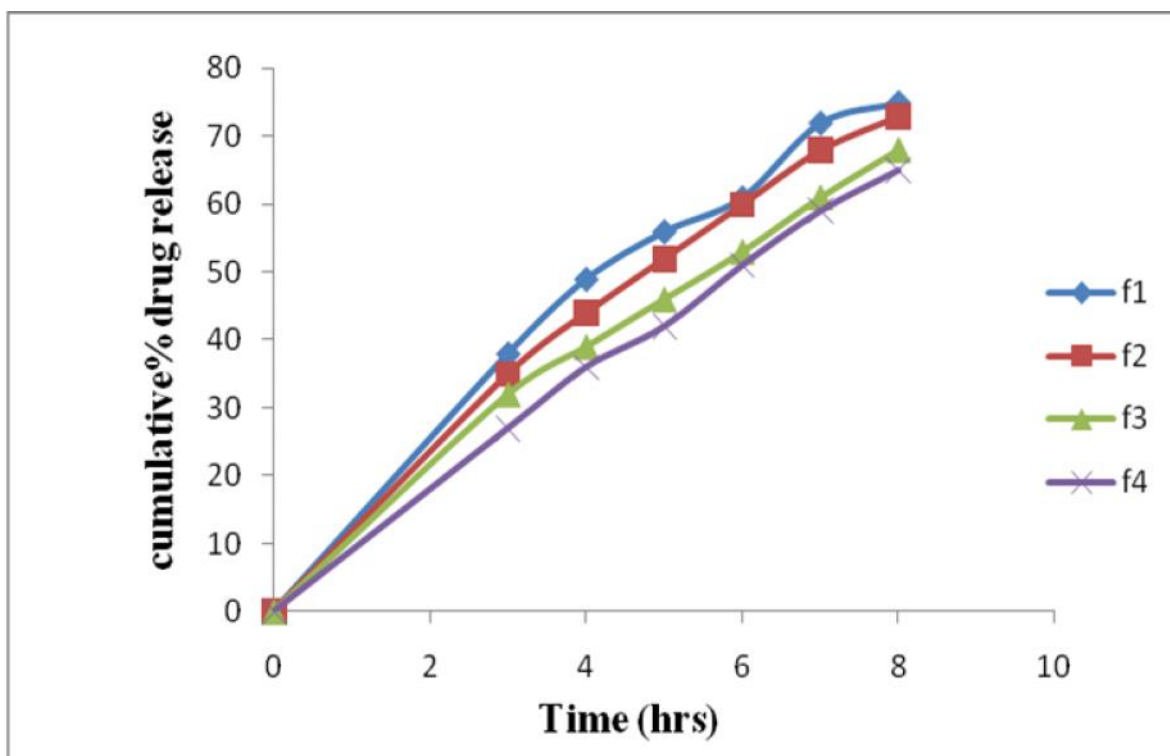


Fig.6 *In vitro* cumulative% drug release of Acyclovir *In situ* gels with xanthan gum in p<sup>H</sup>6.8

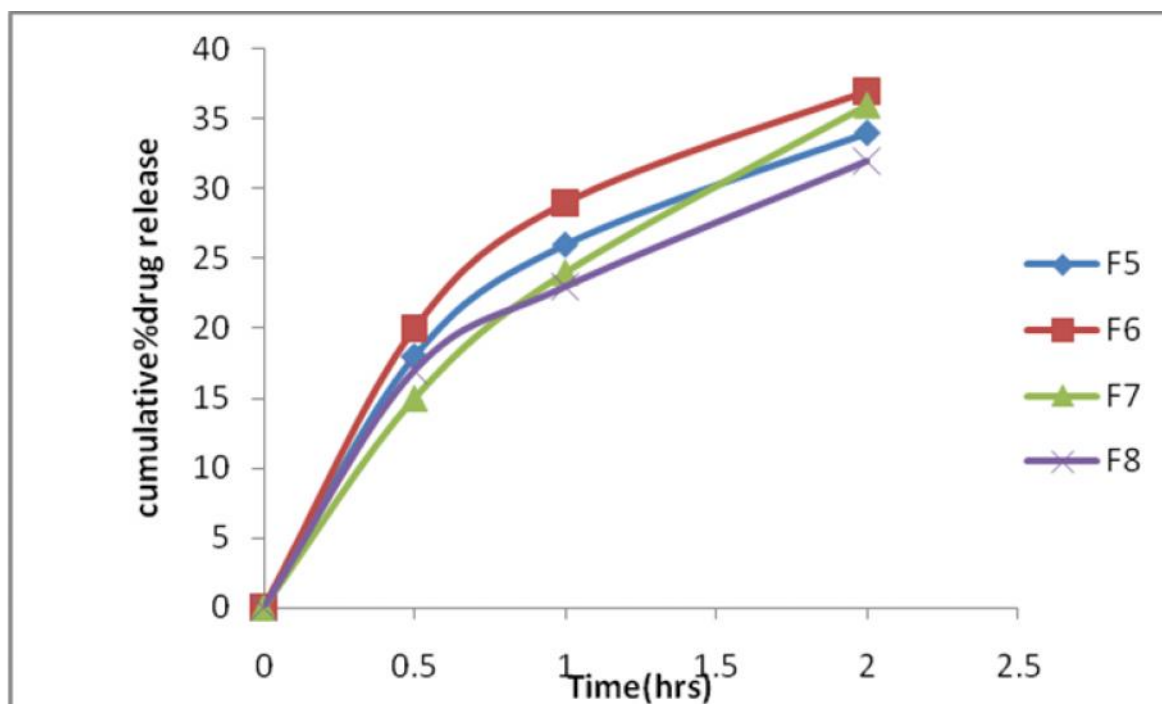
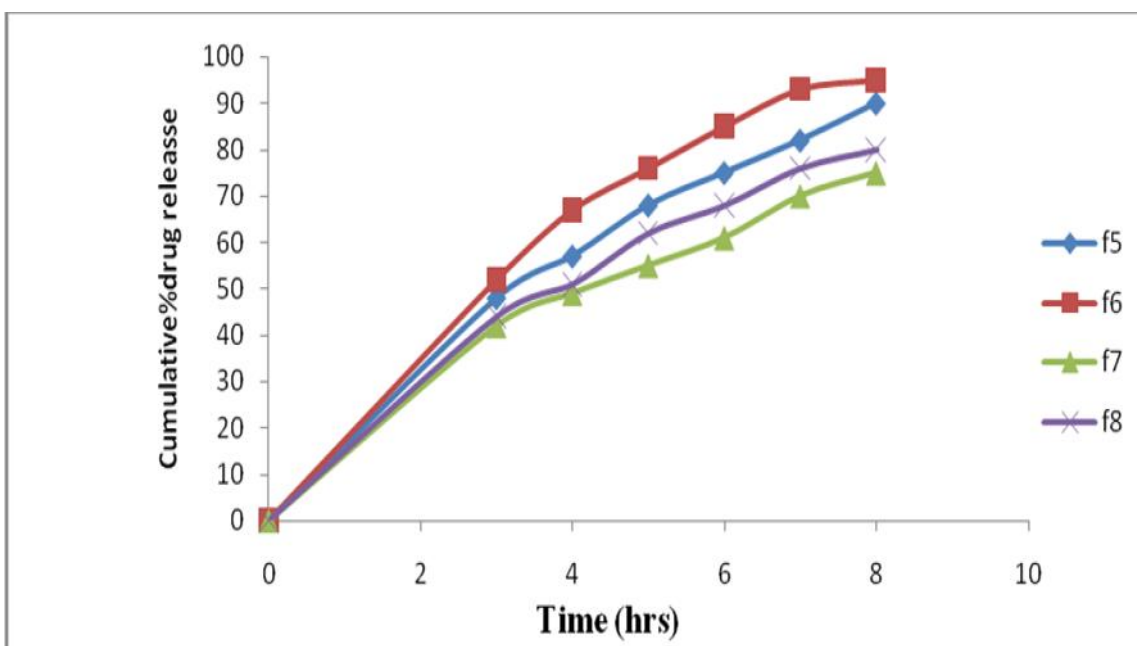


Fig. 7 *In vitro* cumulative% drug release of Acyclovir *In situ* gels with sodium alginate in p<sup>H</sup>1.2





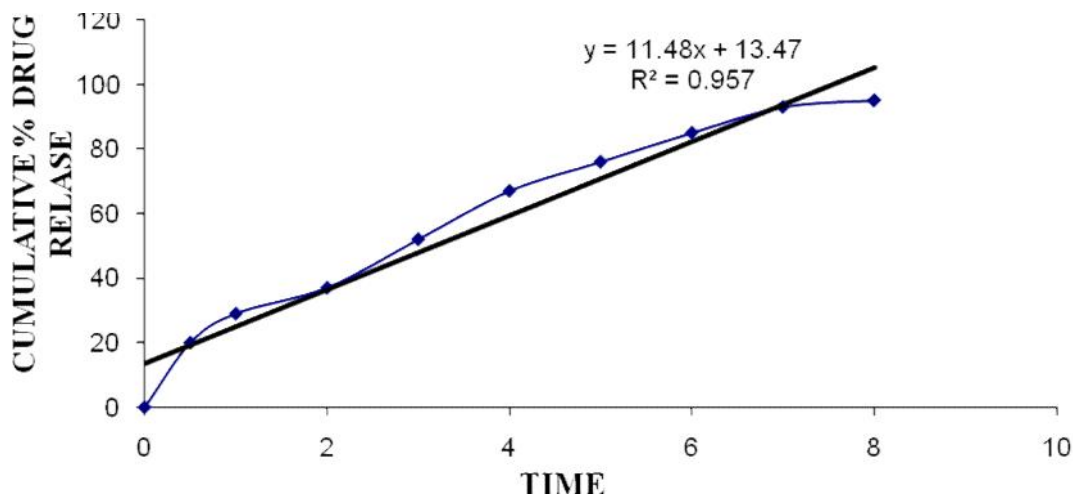
**Fig. 8** *In vitro* cumulative% drug release of Acyclovir *In situ* gels with sodium alginate in pH 6.8

The *In-vitro* drug release study was performed using dissolution rate test apparatus in phosphate buffer (pH 6.8). The dissolution profiles of acyclovir are given in Fig.5-8.

The *In-vitro* cumulative percentage drug release was observed in the range of 95 to 65 percent. The formulations F1, F2, F3, F4, F5, F6, F7, F8 containing

xanthan gum and sodium alginate respectively showed a release of 75, 73, 68, 65,90,95,75 and 80% after 8 hours. This shows that more sustained release was observed with the increase in percentage of xanthan gum and sodium alginate. The best formulation was observed as F6, by the observation of all results of the eight formulations acyclovir *In situ* gels.

**IN VITRO DRUG RELEASE KINETICS**



**Fig.9** Zero order release kinetics of optimized formulation F6

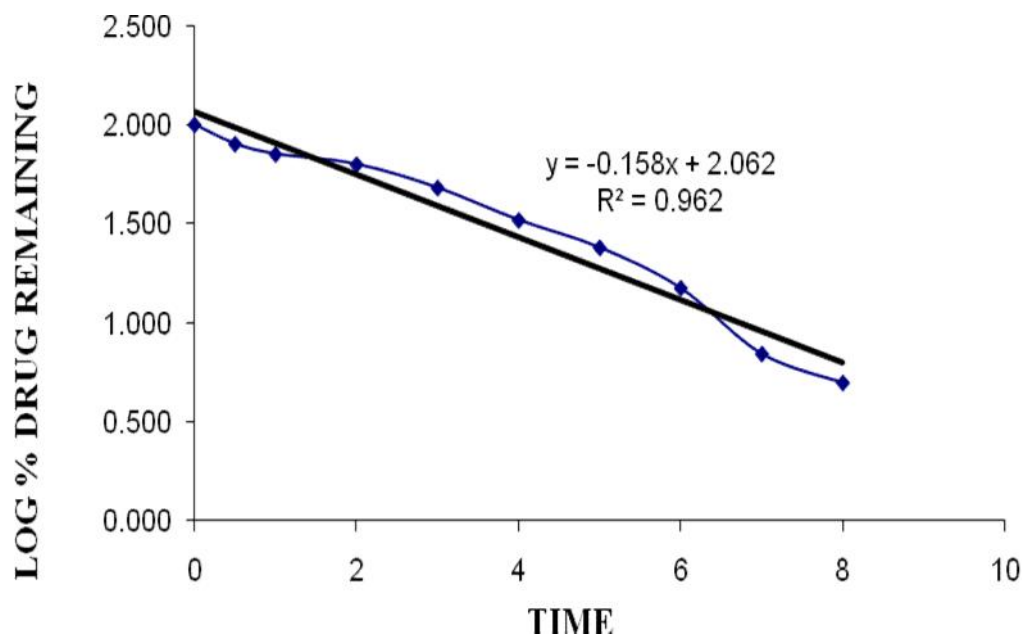


Fig .10 First order release kinetics of optimized formulation F6

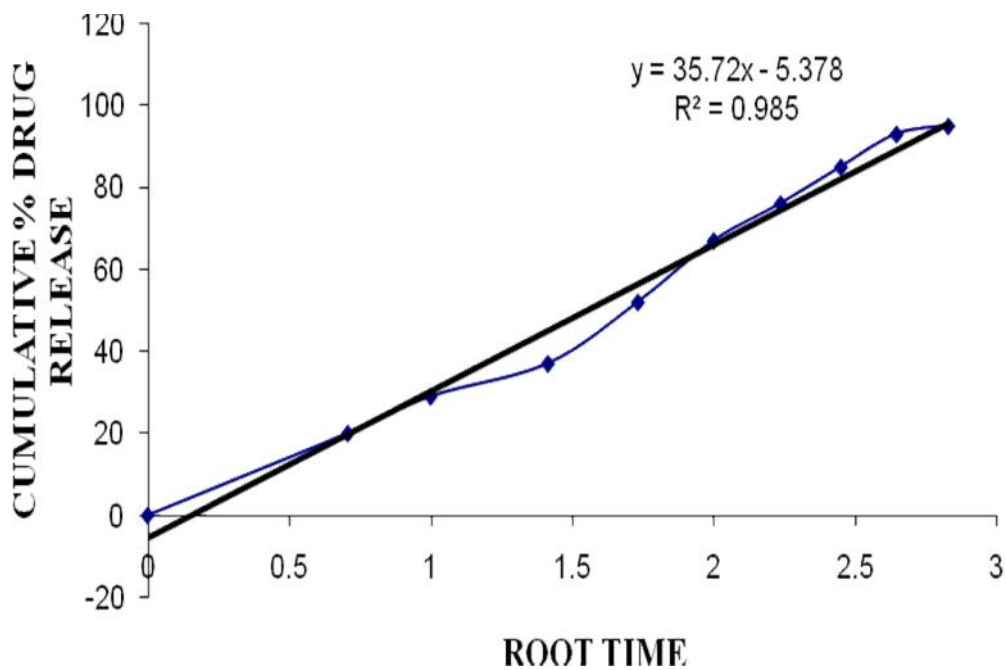


Fig .11 Higuchi release kinetics of optimized formulation F6

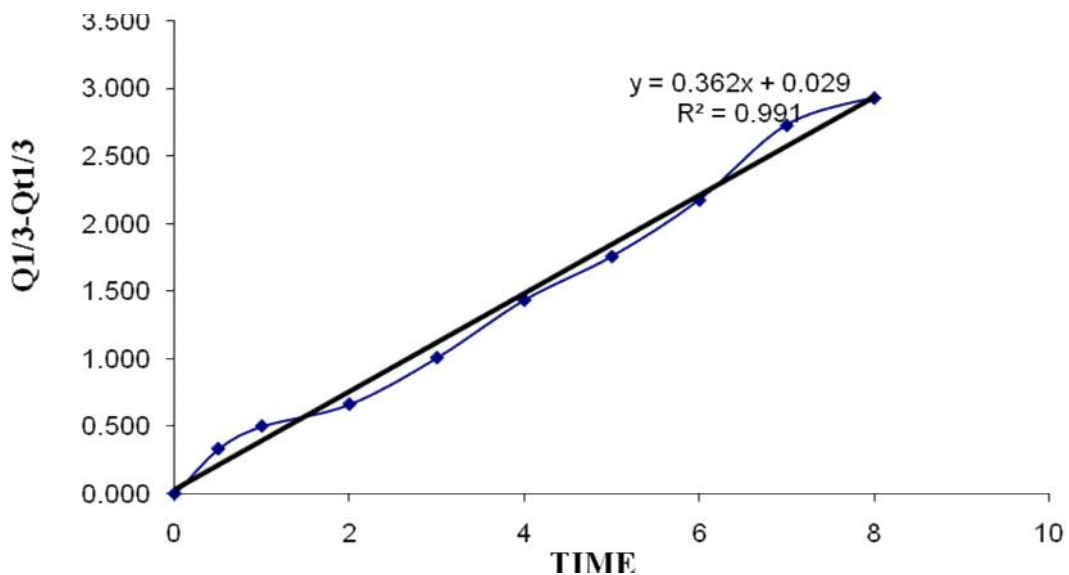


Fig .13 Hixson Crowell release kinetics of optimized formulation F6

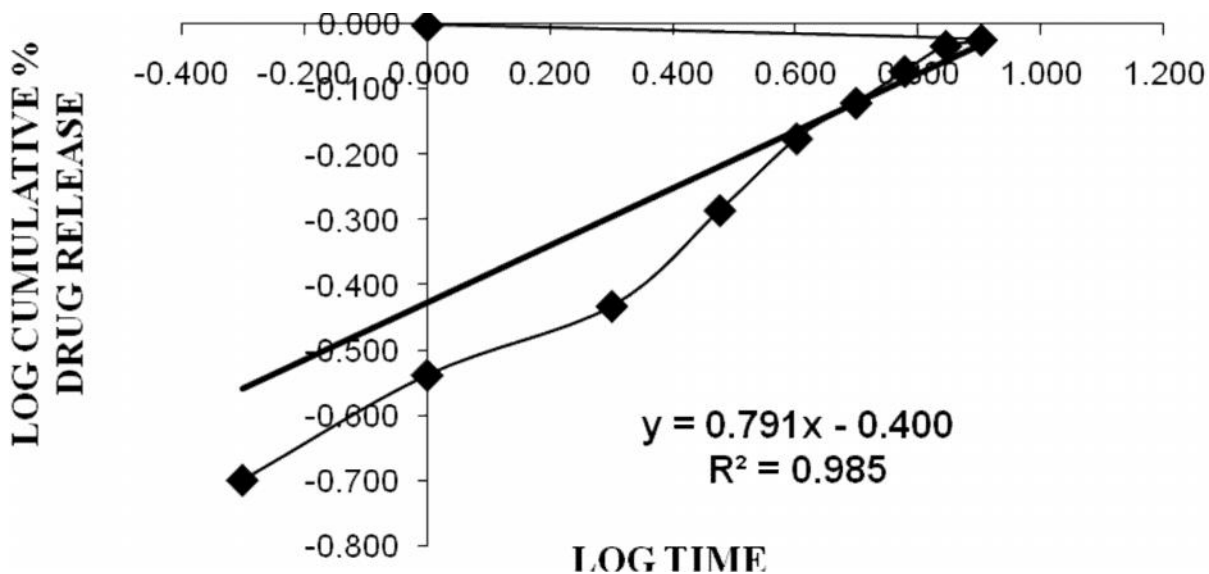


Fig .14 Korse meyer peppas release kinetics of optimized formulation F6

**r<sup>2</sup> values of different kinetic models of *In situ* gels**

Table 6. r<sup>2</sup> values of different kinetic models of optimized formulations F6

| Formulation | Zero order | First order | Higuchi model | Korsemeyer peppas model |       | Hixson Crowell model |
|-------------|------------|-------------|---------------|-------------------------|-------|----------------------|
|             |            |             |               | r <sup>2</sup>          | n     |                      |
| F6          | 0.9577     | 0.8724      | 0.9853        | 0.9822                  | 0.588 | 0.9911               |

Where, r<sup>2</sup> = regression coefficient  
n = slope

Based on the mathematical data revealed from kinetic models, it was concluded that the release data of F6 formulation was best fitted with Hixson Crowell model. The diffusion exponent 'n' values were found to be more than 0.5 indicating non-Fickian diffusion.

## Conclusion

In the present research work we have prepared oral *in situ* gels of acyclovir. Under the preformulation studies, API (Active Pharmaceutical Ingredient) characterization and drug-excipient compatibility studies were carried out. The API characterization showed compliance with the drug characteristics. This study was demonstrated that *In situ* gels formed by oral administration of solutions of sodium alginate and that release of Acyclovir is sustained over a period of at least 8 hrs when compared to xanthan gum.

## Acknowledgments

I express my heartfelt thanks to Mr. K. Sudhir, the honorable Chairman and Mr. T. Raghava Reddy, honourable secretary, St, Pauls group of colleges, for providing the facilities to finish the project in our college.

## References

- 1 Koelle DM, Corey L (2008). "Herpes simplex: insights on pathogenesis and possible vaccines". *Annual Review of Medicine* 59: 381–95.
2. Miyazaki, N. Kawasaki, K. Endo, D. Attwood. *In situ* gelling gellan formulations as vehicles for oral drug delivery. *J. Control Rel.* 1999; 60:287-95.
3. Miyazaki S, Suzuki, N. Kawasaki, K. Endo, A. Takahashi, D., Oral sustained delivery of paracetamol from *in situ* gelling xyloglucan formulations. *Drug Dev Ind. Pharm.* 2003; 29 (2):113-9.
4. Peppas N and Langer R. New challenges in biomaterials. *Science* 1994; 263:1715-20.
5. A. Rozier, C. Mazuel, J. Grove, B. Plazonnet, Gelrite,: A novel, ionactivated, *in situ* gelling polymer for ophthalmic vehicles. Effect on bioavailability of timolol. *Int. J. Pharm.* 1989; 57 163–168.
6. S. Cohen, E. Lobel, A. Trevigoda, Y. Peled.. A novel *In situ*-forming ophthalmic drug delivery system from alginates undergoing gelation in the eye. *J. Control. Release.* 1997; 44 201–208.
7. B. Srividya, R.M. Cardoza, P.D. Amin. . Sustained ophthalmic delivery of ofloxacin from a pH triggered *in situ* gelling system. *J. Control Release.* 2001;73: 205–211.
8. S. Miyazaki, N. Kawasaki, K. Endo, D. Attwood. Oral sustained delivery of theophylline from thermally reversible xyloglucan gels in rabbits. *J. Pharm. Pharmacol.* 2001; 53: 1185–1191.

9. S. Miyazaki, S. Suzuki, N. Kawasaki, K. Endo, A. Takahashi, D. Attwood. *In situ* gelling xyloglucan formulations for sustained release ocular delivery of pilocarpine hydrochloride. *Int. J. Pharm.* 2001; 229: 29–36.
10. S. Miyazaki, F. Suisha, N. Kawasaki, M. Shirakawa, K. Yamatoya, D. Attwood.. Thermally reversible xyloglucan gels as vehicles for rectal drug delivery. *J. Control. Release.* 1998; 56: 75–83.
11. Himanshu Gupta and Aarti Sharma. Ion activated bioadhesive *in situ* gel of clindamycin for vaginal application. *International Journal of Drug Delivery.* 2009; 1: 32-40.
12. U.V. Singh, N. Udupa, R. Kamath, P. Umadevi,, Enhanced Biodegradable *in situ* forming implants and methods of antitumor efficacy of methotrexate poly(lactic-co-glycolic) producing the same, US Pat. 4938763, 3 July 1990. acid injectable gel implants in mice bearing sarcoma-180, 136.
13. F. Suisha, N. Kawasaki, S. Miyazaki, M. Shirakawa, K. Yamatoya, M. Sasaki, D. Attwood, Xyloglucan gels as sustained release vehicles for the intra peritoneal administration of mitomycin C. *Int. J. Pharm.* 1998 ; 172: 27–32.
14. [wikipedia.org/wiki/Aciclovir](http://wikipedia.org/wiki/Aciclovir).